Isolation of Mouse Embryonic Fibroblasts
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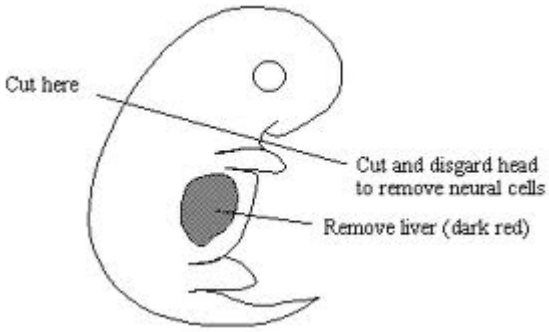
Procedure:

1) Collect embryos at E13.5 or E14.5
    a. Wash embryos in phosphate buffered saline (PBS)
2) Using forceps, remove placenta and membranes from embryo. Dissect out liver using forceps and cut off head using scalpel (see image). Place embryos in a clean petri dish.



3) Finely mince tissue using forceps. Add 10 ml of 0.05% Trypsin-EDTA (Gibco, 25300-054) and continue to mince tissue by pipetting tissue up and down. Incubate tissue at 37 °C for 20 minutes.
4) Resuspend tissue and re-incubate at 37 °C for an additional 15 minutes.
5) Neutralize the Trypsin-EDTA digest solution by adding 20 ml of culture medium (DMEM + 10 % FBS + antibiotics) and transfer contents through 70 μm strainer into 50 ml conical tube.
6) Pellet cells via centrifugation at 1 kRPM for 5 min
7) Aspirate media and add 20 ml of culture medium. Plate onto a T25 flask (1 embryo/flask). Place flask in 37 °C incubator overnight.
8) Change medium to get rid of dead cells and cell debris.
9) Can freeze cells when at 80-90 % confluency or can use cells for experiments.